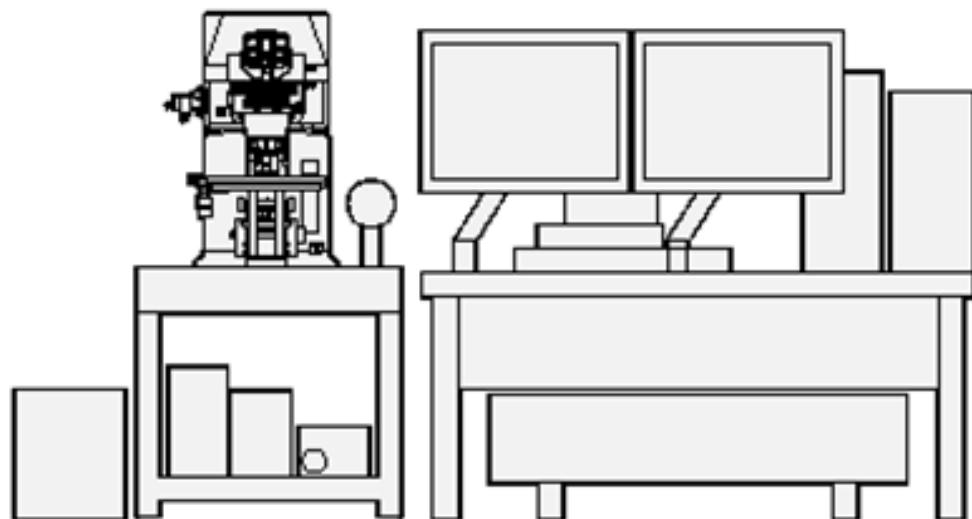


**OLYMPUS®**

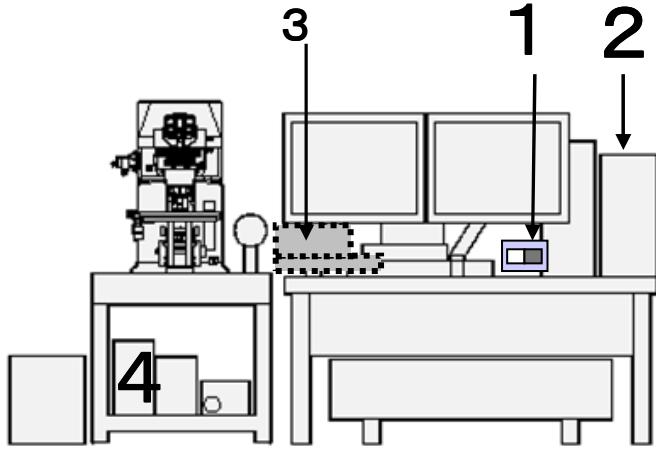
**Confocal Laser Scanning Microscopy**  
**FV1000-D**  
**BX61 (Filter Type)**  
**Operation Manual**



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# Starting the System



3



6



1. Turn the Concentrated Power Supply ON.

2. Turn the computer ON.

3. Turn the laser ON.

HeNe-G(543nm) : Black power supply.

Turn the key switch ON.

(473nm is connected to concentrated power supply)

4. Turn the mercury burner ON for Fluorescence observation.

5. Log on Windows

User ID: [fluoview](#)

Password: [fluoview](#)

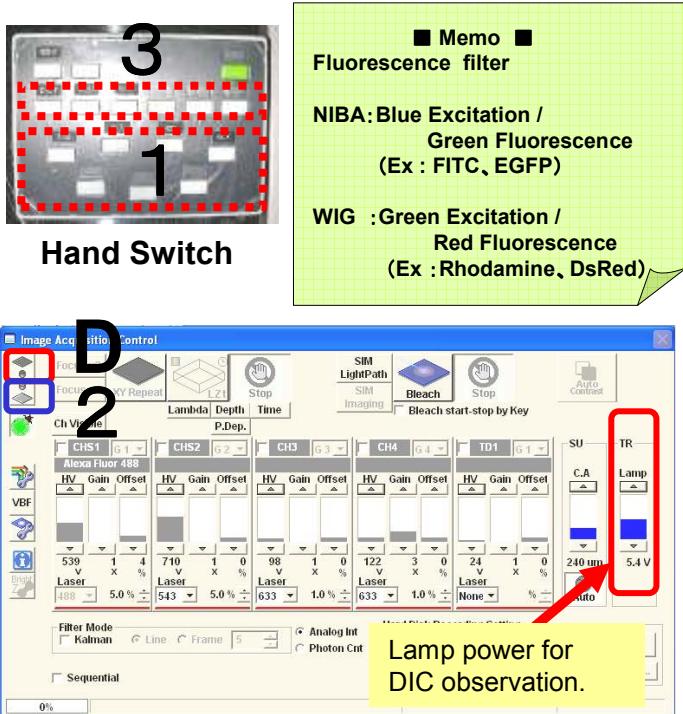
6.  Double click this icon to log on to FV10-ASW

User ID: [Administrator](#)

Password: [Administrator](#)

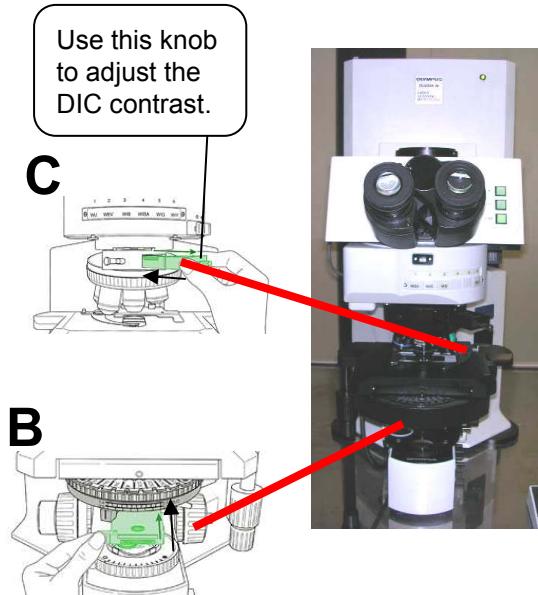
# Visual Observation under the Microscope

## ■ ■ Observation of Fluorescence Image ■ ■



1. Select an objective lens by using the hand switch.
2. Click the button on the Fluoview software.
3. Select fluorescent filter cube by using the hand switch.
4. Focus to the specimen.
5. After observation, click the button to close the shutter.

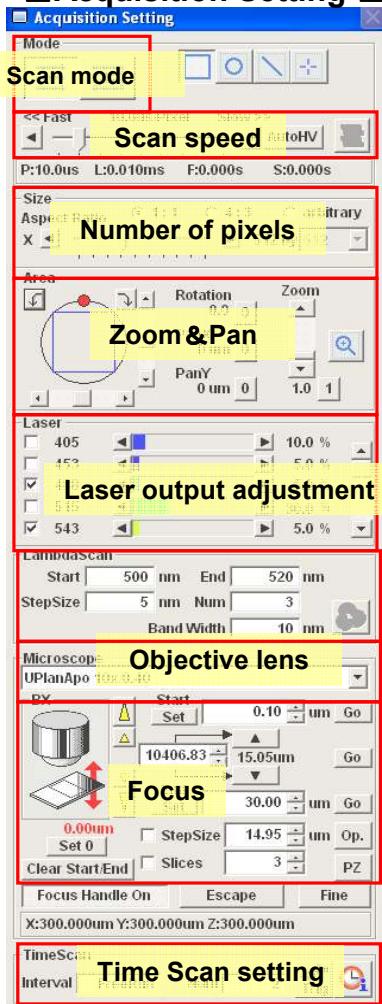
## ■ ■ Observation of Differential Interference Contrast Images ■ ■



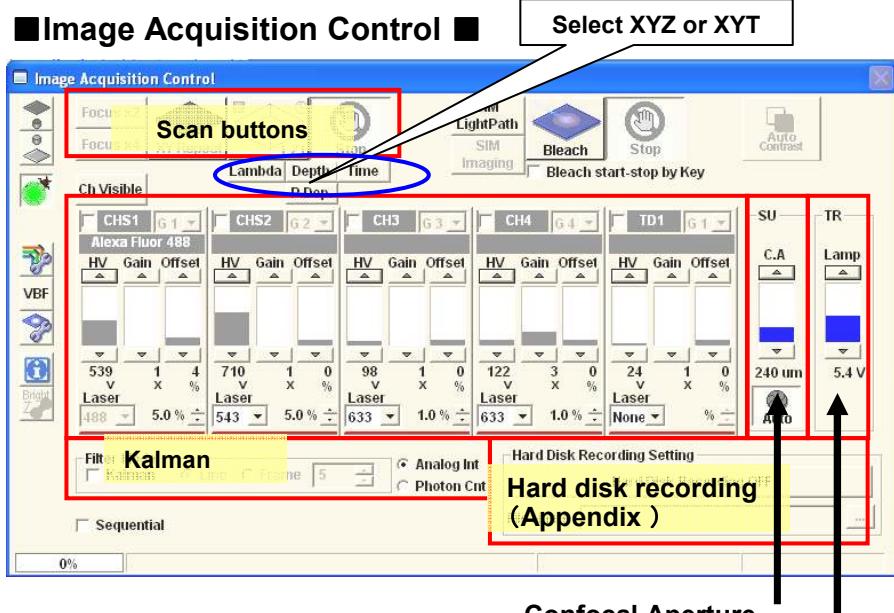
- A. Select the Objective Lens.
- B. Insert the Polarizing Plate in the Light Pass.
- C. Insert the DIC prism slider in the Light Pass.
- D. Click the button on Fluoview software.
- E. Focus to the specimen.
- F. After observation, click the button to close the shutter.

# Overview of Operation Panel for Image Acquisition

## ■ Acquisition Setting ■

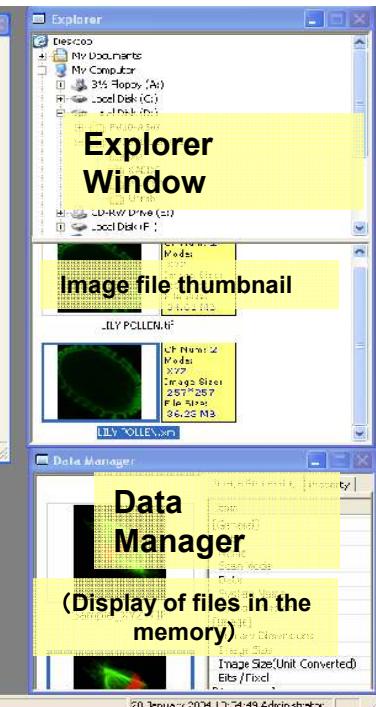
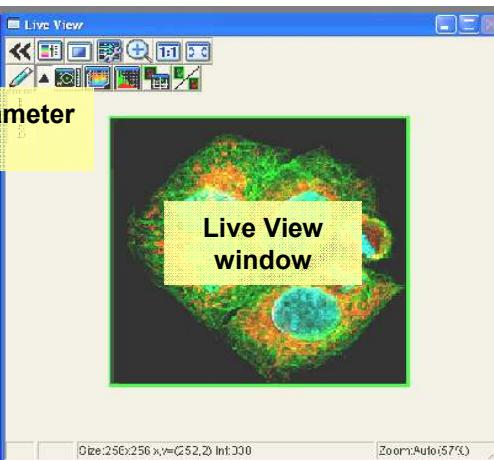
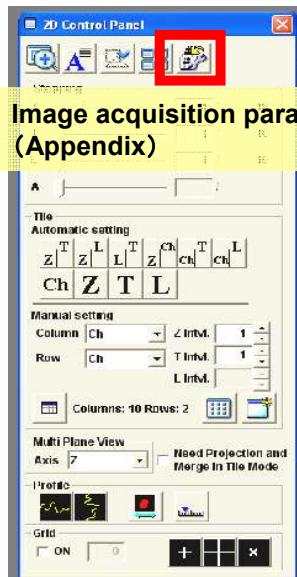


## ■ Image Acquisition Control ■



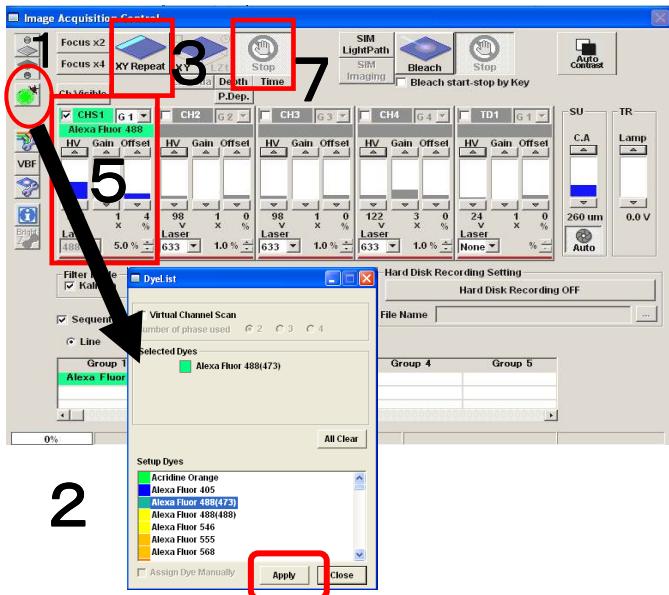
Confocal Aperture

Light intensity  
Adjustment for  
Halogen bulb

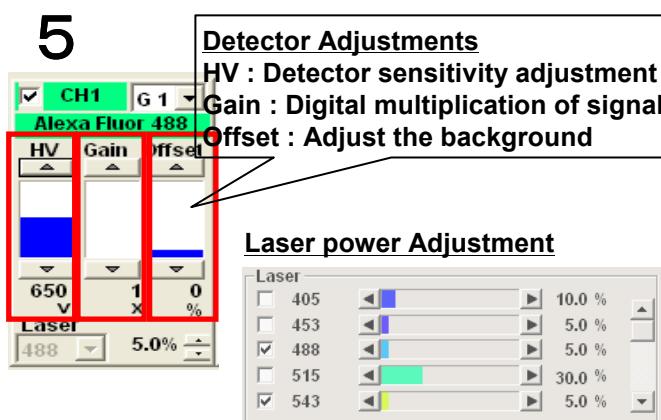


## Image Acquisition (Single Stain on XY Image)

## Sample : Single stain of green fluorescence dye (Alexa fluor 488)



5



## 8 Rotation

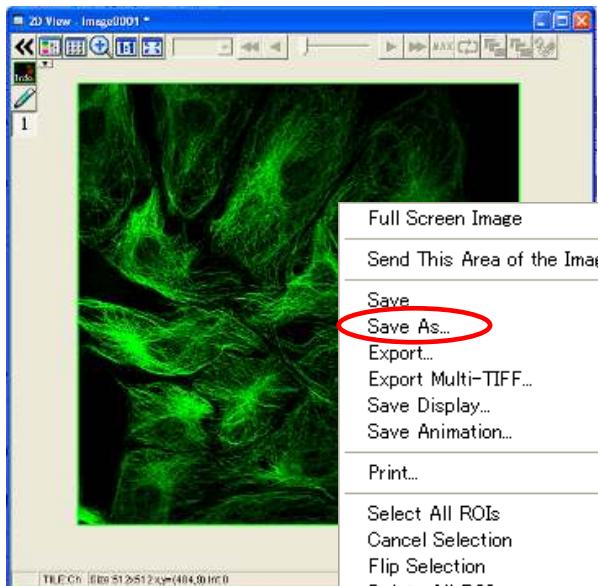
1. Click on the ‘Dye List’ button. On the Dye List panel, double-click on a fluorescence reagent to be used for observation.
2. Click on the Apply button.
3. Click XY Repeat button  to get live image.
4. Focus to the specimen.
5. Adjust the green (Alexa488) image.  
(Laser power, HV, Offset, etc)
6. Use zoom and rotation if it is needed.
7. After the image adjustments, click Stop button  to stop scanning.
8. Select AutoHV and then select Scan speed.

※As the scan speed becomes slower, noise can be removed while maintaining the current brightness.  
You can also use Kalman, to remove the noise. Look at P11.

# Image Acquisition (Single Stain on XY Image)

Sample : Single stain of green fluorescence dye (Alexa fluor 488)

9



9. Click XY button to Acquire the image.

10. Saving the image : Right-click on the 2D view and select Save As to save the image.

## ■ Memo ■ File formats specifically for the FV10-ASW

OIF format : Creates “a folder that contains an image (16-bit TIFF)” and “an accessory file,” which cannot be opened separately from each other.

OIB format : Creates the OIF format files in a single file, which is convenient for migration and other operations.

Look at Appendix1 to export the images to TIFF, BMP, or JPEG.

## ■Adjustment brightness intensity■

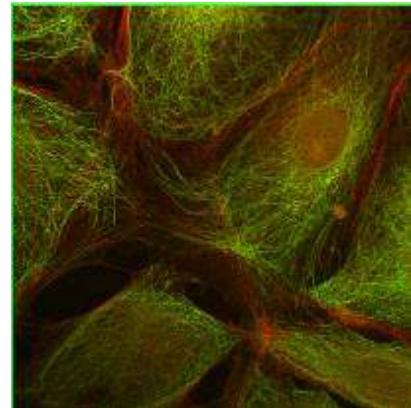
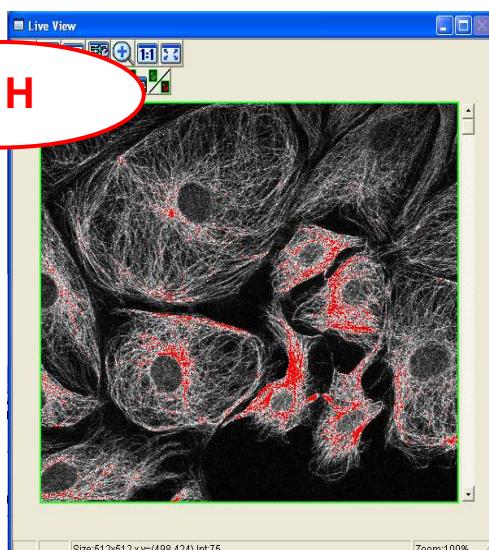
Press keyboard Ctrl + H key

Optimized PMT adjustment brightness intensity 2 color between white and black, Maximum intensity is 4095(12bit).

If intensity is over4095, color is changed to red (saturation).

If intensity is under 0, color is changed to blue (no signal).

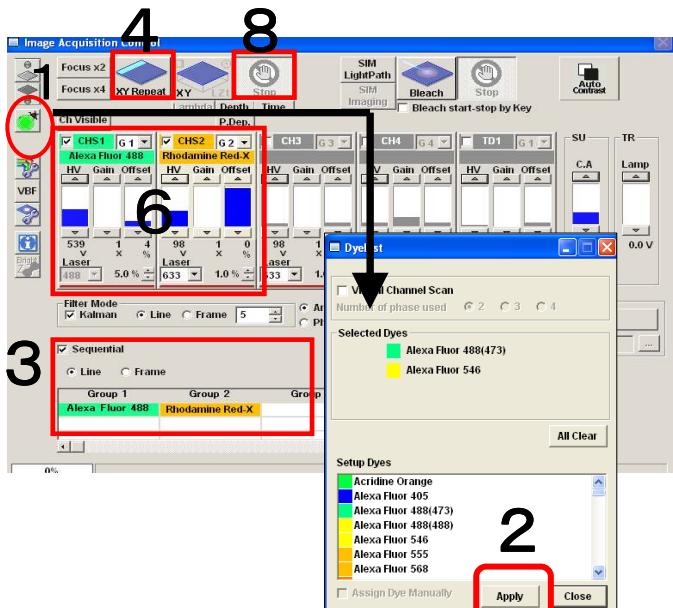
Ctrl+H



## Image Acquisition (Double stain on XY)

## Sequential scan

※ Sequential scan is a one way to remove the fluorescence cross talk.



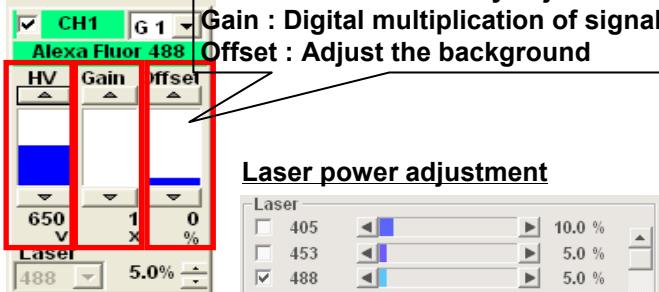
2

6

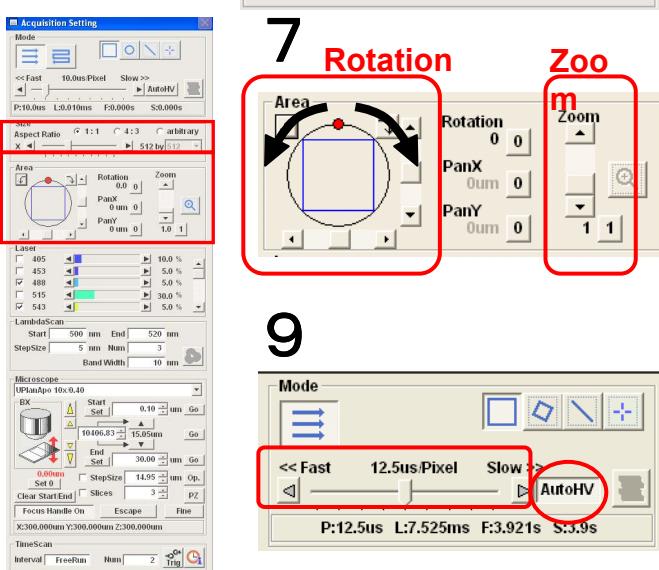
## Detector Adjustments

## HV : Detector sensitivity adjustment

## Gain : Digital multiplication of signals



## Laser power adjustment



9

## Rotation

**Zoo**

1. Click on the 'Dye List' button. On the Dye List panel, double-click on a fluorescence reagent to be used for observation.

2. Click on the Apply button.

3. Check 'Sequential', and select 'Line'.

4Click XY Repeat button  to get live image.

## 5. Focus to the specimen.

6. Adjust the green (Alexa488) image.  
( Laser power, HV, Offset, etc )

7. Use zoom and rotation if it is needed.

8. After the image adjustments, click Stop button  to stop scanning.

9. Select AutoHV and then select Scan speed.

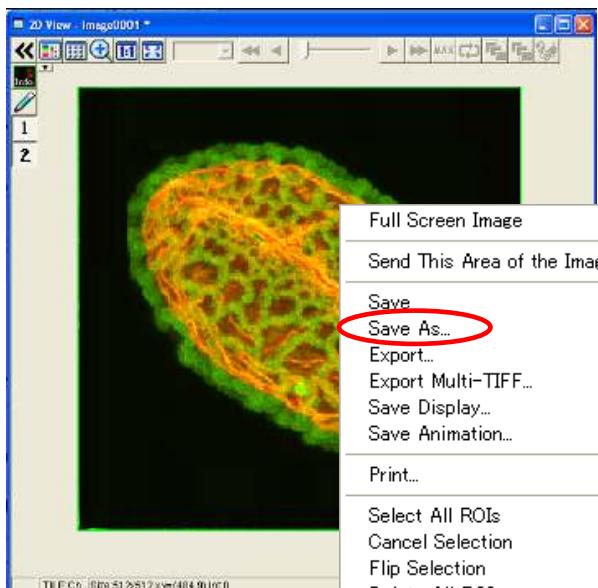
※As the scan speed becomes slower, noise can be removed while maintaining the current brightness.

You can also use Kalman, to remove the noise. Look at P11.

# Image Acquisition (Double stain on XY)

## Sequential scan

10



9. Click XY button to Acquire the image.

10. Saving the image : Right-click on the 2D view and select **Save As** to save the image.

### ■ Memo ■ File formats specifically for the FV10-ASW

OIF format : Creates “a folder that contains an image (16-bit TIFF)” and “an accessory file,” which cannot be opened separately from each other.

OIB format : Creates the OIF format files in a single file, which is convenient for migration and other operations.

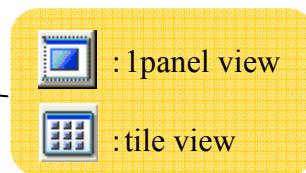
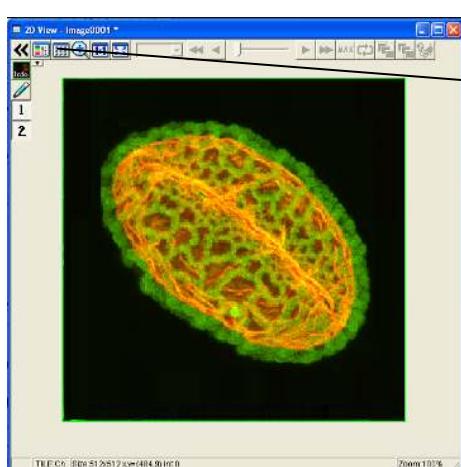
Look at Appendix1 to export the images to TIFF, BMP, or JPEG.

## ■ Change View mode ■

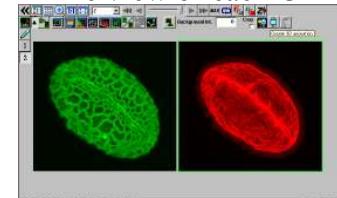
In Live View and 2DView, there are three pattern for multicolor sample.

1. 1 panel of merge image.
2. Tile view of each channel image.
3. Tile view of each channel and merge image.

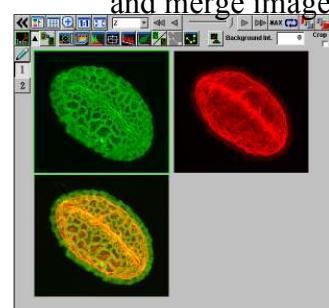
Tile view and 1 panel view is changed on the icons and if you need each channel and merge channel, select ‘Add a View’ on the right click menu.



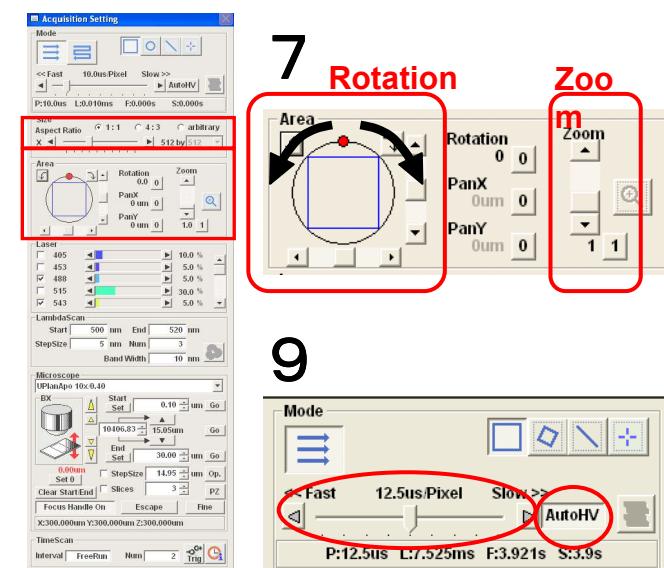
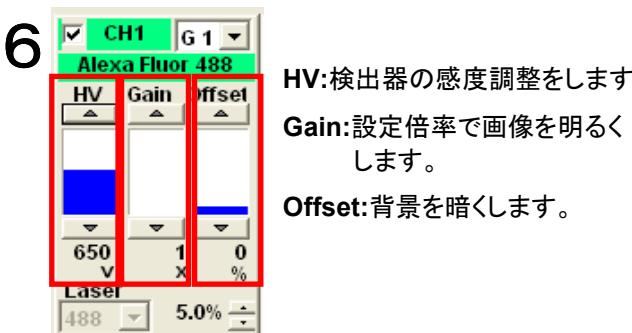
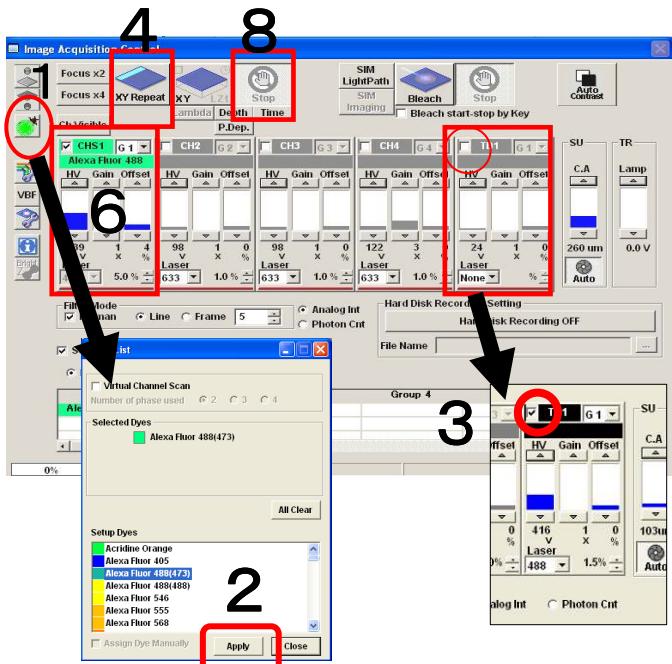
2. The view of each CH



3. The view of each CH and merge image



# Image Acquisition (DIC with fluorescence on XY)



1. Click on the 'Dye List' button. On the Dye List panel, double-click on a fluorescence reagent to be used for observation.

2. Click on the Apply button.

3. Check 'TD1'.

4. Click XY Repeat button to get live image.

5. Focus to the specimen.

6. Adjust the green (Alexa488) image. (Laser power, HV, Offset, etc)

7. Use zoom and rotation if it is needed.

8. After the image adjustments, click Stop button to stop scanning.

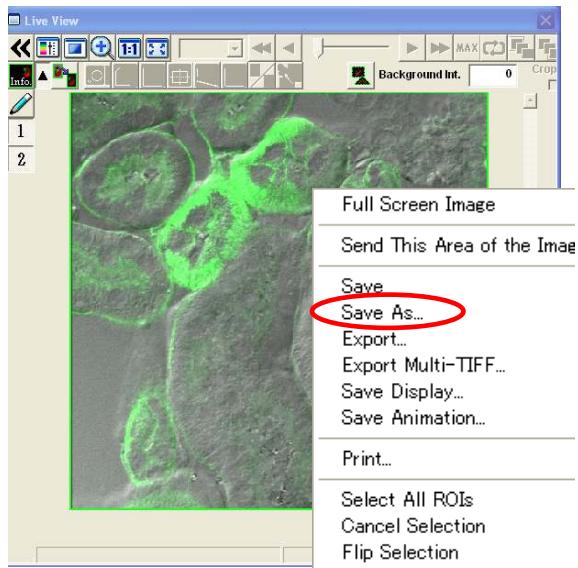
9. Select AutoHV and then select Scan speed.

※As the scan speed becomes slower, noise can be removed while maintaining the current brightness.

You can also use Kalman, to remove the noise. Look at P11.

# Image Acquisition (DIC with fluorescence on XY)

9



9. Click XY button to Acquire the image.

10. Saving the image : Right-click on the 2D view and select **Save As** to save the image.

## ■ Memo ■ File formats specifically for the FV10-ASW

OIF format : Creates “a folder that contains an image (16-bit TIFF)” and “an accessory file,” which cannot be opened separately from each other.

OIB format : Creates the OIF format files in a single file, which is convenient for migration and other operations.

Look at Appendix1 to export the images to TIFF, BMP, or JPEG.

## 【Points to get a good images】

### Signal is too dark

**Turn up HV**: Turn up sensitivity of detector. If you cannot get a good brightness in over 700V, take another methods.

**Turn up Laser power**: Fluorescence brightness will raise by laser power. But it is too much power, bleach occurs.

**Changing CA**: In normally, button is on to set the perfect size of pinhole by automatically. If you want to change it, undo the button and change it.

**Turn up Gain**: Digital multiplication of signal.

### Too many noise

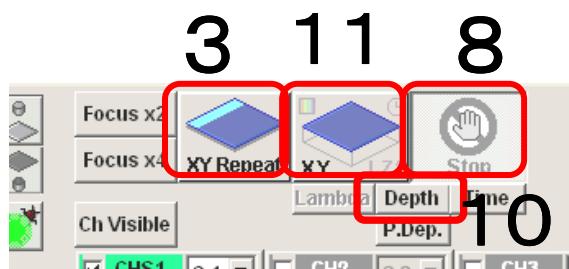
**Kalman**... Average the images to remove the random noise.

Click Kalman, and select Line and input the number of images for average.



# Image Acquisition (XYZ)

## Acquisition of 3D image



1. Adjust XY image.

Look at from page 6 to page 9

2. Click 'Clear Start/End' to reset the upper and lower position. And also, click 'Set 0' to change the base point.

3. Click XY Repeat button to get live image.

4. Click this to change the Z position and decide upper position.

5. Click Start 'Set' button to set the upper position.

6. Click this to change the Z position and decide lower position.

7. Click End 'Set' button to set the lower position.

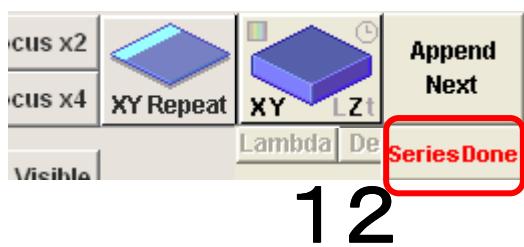
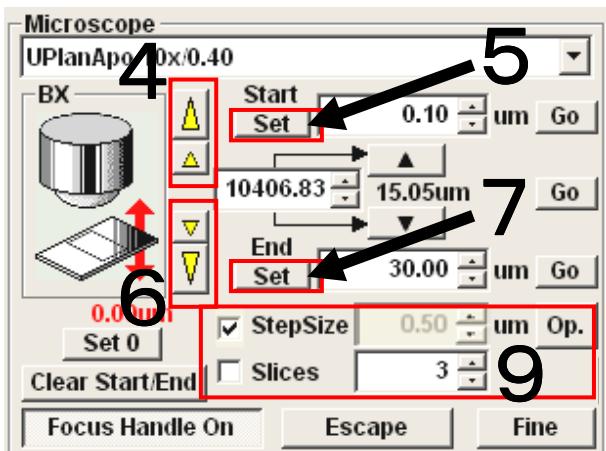
8. Click Stop button to stop live view.

9. Put in a value to StepSize or Slices.  
(You can see the recommendation by Op. button)

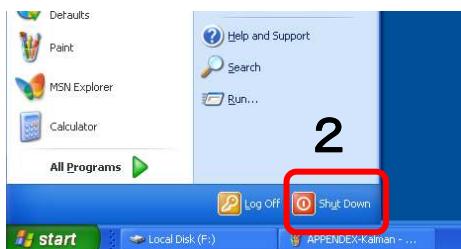
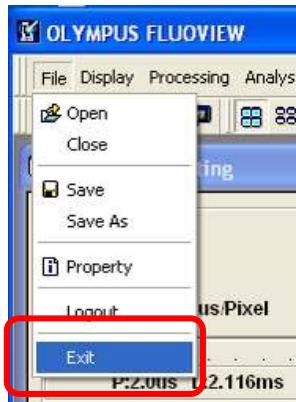
10. Click on 'Depth'.

11. Click XYZ button to acquire the XYZ image series.

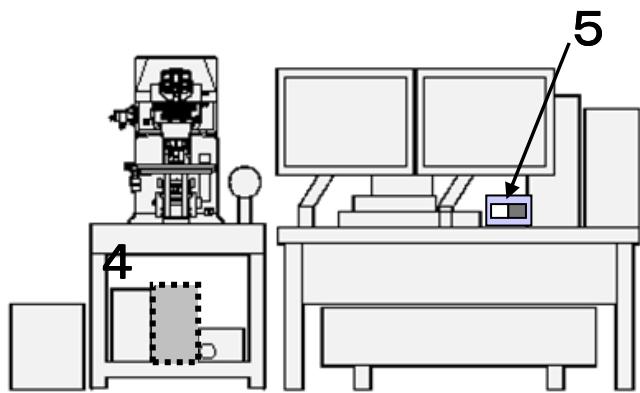
12. Click 'Series Done' and save the image.



# Closing the System



3



1. File/Exit to shut down the software.

2. Shutdown Windows.

3. Turn laser off.

**HeNe-Green laser (543nm)**

4. Turn the mercury burner Off.

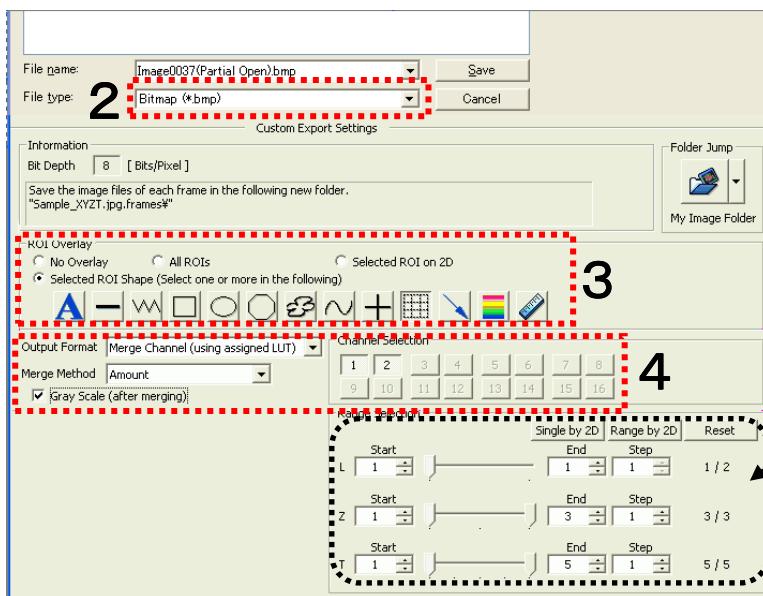
5. Turn the Concentrated Power Supply Off.

## Appendix 1

# Export Images

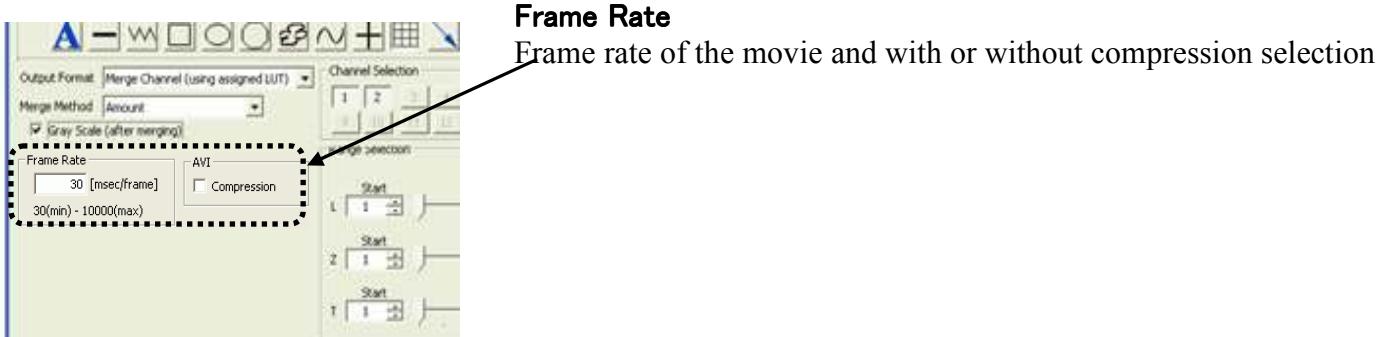
### Export to Bitmap (\*.bmp) · JPEG (\*.jpg, \*.jpeg) · TIFF (\*.tif, \*.tiff) · PNG (\*.png)

1. Choose 'Export' in the right click menu on the 2D View window of data to export.
2. Select file directory, name and format.
3. Select ROI shape if you needed. (ex scale)
4. Choose format in Output Format, channels in Channel Selection and range in Range Selection (Z/T).
5. Click 'Save' to export.



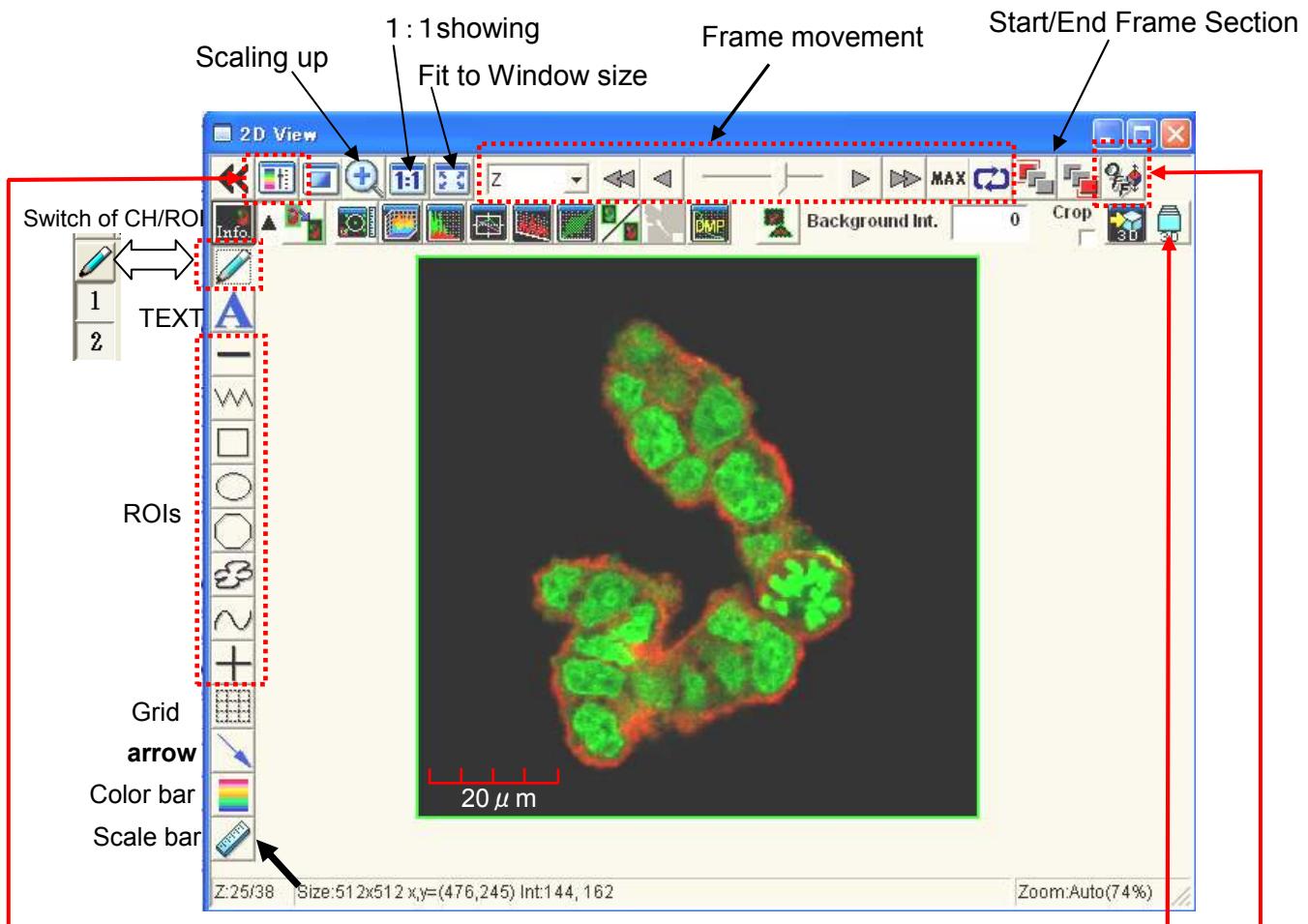
### Export to Movie Files (AVI, MOV形式は、XYZ・XYT・XYλ・XZT・XλT・XλZ画像で可能です)

1. Choose 'Export' in the right click menu on the 2D View window of data to export.
2. Select file directory, name, format, ROI shape, Output format, Channels and range.
3. Input the Frame Rate.
4. Click 'Save' to export.



## Appendix2

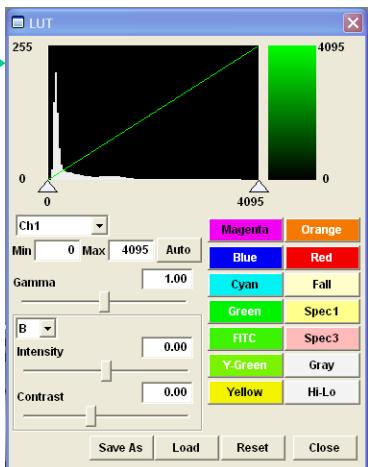
# 2D View Window



Make rotating 3D animation



### LUT settings

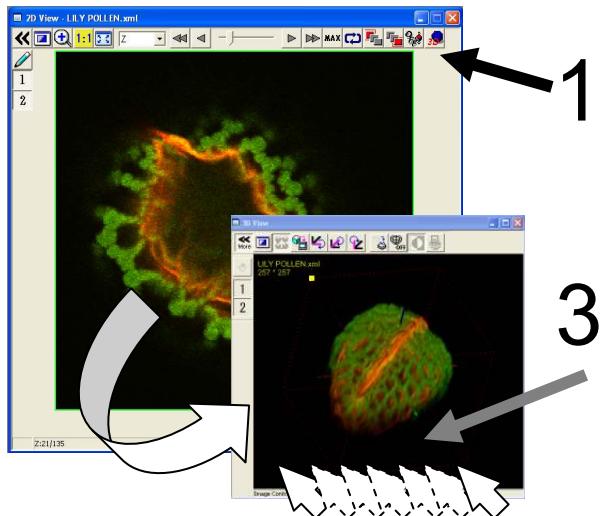


### Projection switching

- (Projection Off) button: no projection.
- (Z intensity projection) button: Z intensity projection is displayed.
- (T intensity projection) button: T intensity projection is displayed.
- (Lambda intensity projection) button: Lambda intensity projection is displayed.
- (Z topographic projection) button: Z topographic projection is displayed.
- (Lambda topographic projection) button: Lambda topographic projection is displayed.
- (T series average) button: The average of T series images is displayed.

## Appendix3

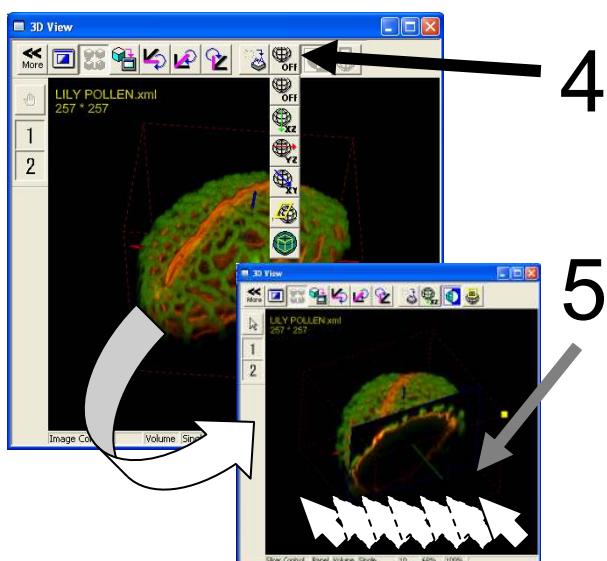
### Image Analysis (Creating a Three-dimensional Image)



Observe the image at a certain angle

1. Click on the  button for a 2D View-(file name) image.
2. A 3D view is created.
3. Drag the mouse on the image to observe it at a certain angle.

→ To save this image, proceed to step 6 on the next page.



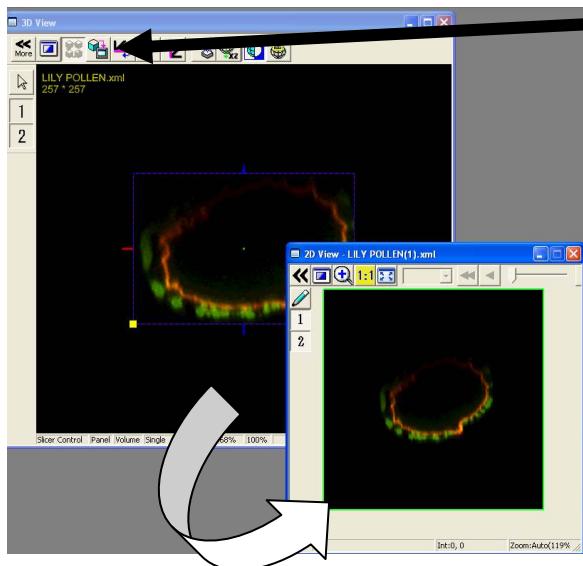
Observe a certain cross-section surface of the image

4. Click on the  button and select .
5. Drag the mouse from side to side on the image to observe a certain vertical section.

→ To save this image, proceed to step 6 on the next page.

## Appendix3

# Image Analysis (Creating a Three-dimensional Image)

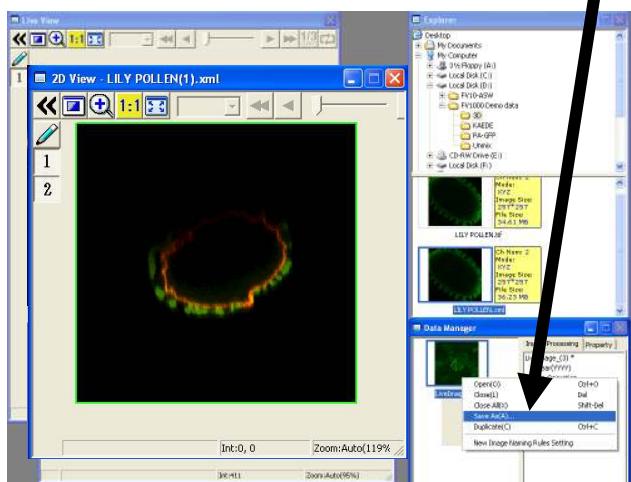


6

Save the image in step 3 or 5



6. Click on the  button.



8

7. A 2D View-(file name) image is created.

8. Saving the image:  
Right-click on the Image Icon displayed on the Data Manager and select Save As to save the image.  
(Save as types "oib" and "oif" are file formats specifically for the FV10-ASW software.)

### ■Memo■

File formats specifically for the FV10-ASW

#### OIF format:

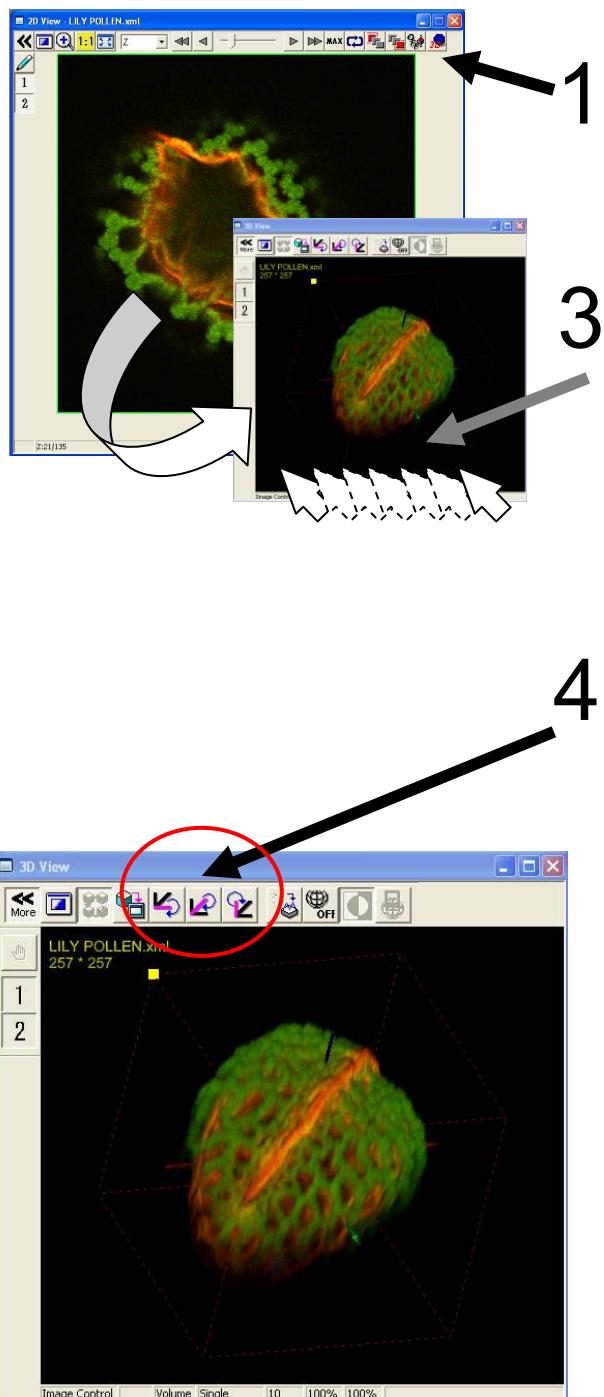
Creates "a folder that contains an image (16-bit TIFF)" and "an accessory file," which cannot be opened separately from each other.

#### OIB format:

Creates the OIF format files in a single file, which is convenient for migration and other operations.

## Appendix4

### Image Analysis (Rotating a Three-dimensional Image)



1. Click on the  button for a 2D View-(file name) image.
2. A 3D view is created.
3. Drag the mouse on the image to observe it at a certain angle.

#### Simple animation

4. Press and hold the  button to rotate the image around the X-axis. Press it again to stop rotation.



Press and hold the  button to rotate the image around the X-axis. Press it again to stop rotation.



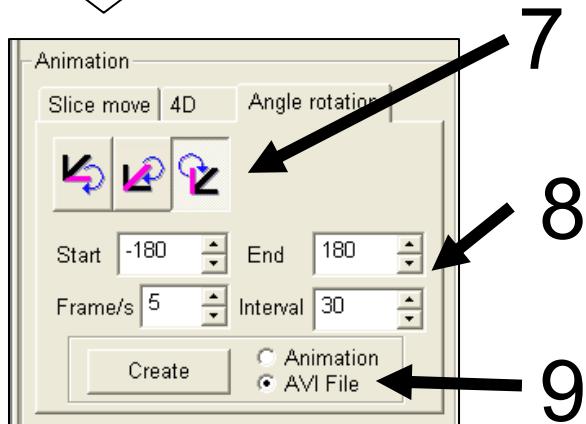
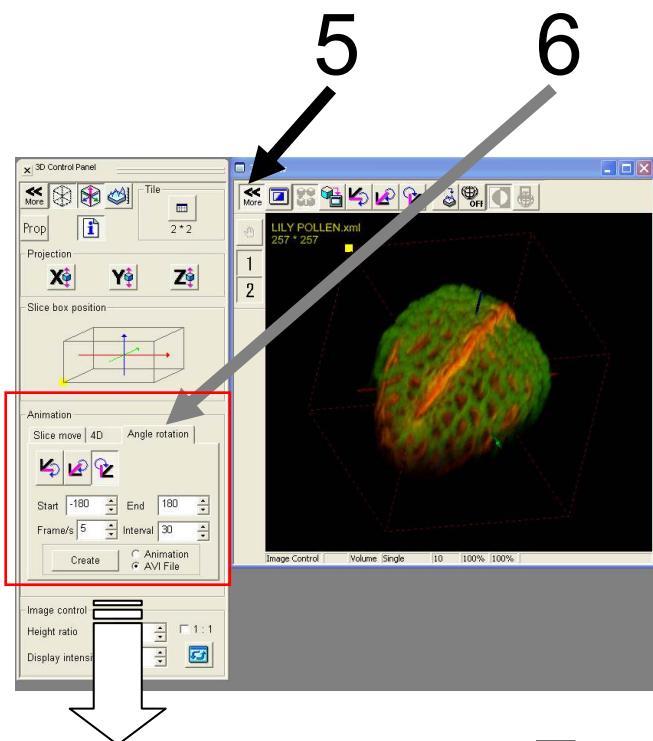
Press and hold the  button to rotate the image around the Y-axis. Press it again to stop rotation.



Press and hold the  button to rotate the image around the Z-axis. Press it again to stop rotation.

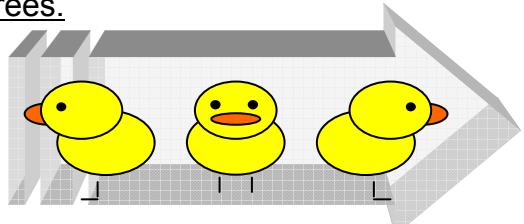
## Appendix4

# Image Analysis (Rotating a Three-dimensional Image)



To save a rotation file as an animated image, create three-dimensional images according to the following procedure.

For example, try to rotate an image by 180 degrees.



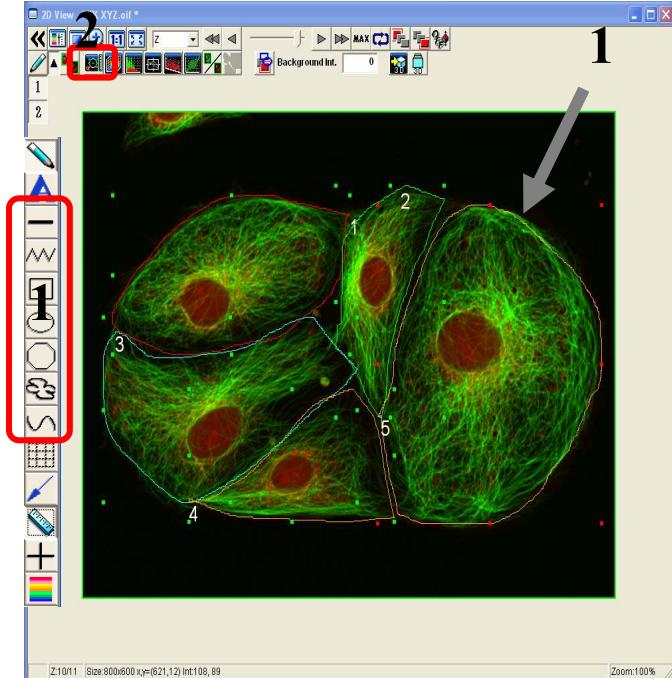
5. Click on the  button.
6. Click on the Angle rotation tab.
7. Select the rotation axis.
8. Enter the rotation angle.

**Start** = Angle to start rotation  
**End** = Angle to stop rotation  
**Frame/s** = Rotation speed  
**Interval** = Degrees to be rotated at a time

9. Select AVI File and click on Create.
10. Enter a file name and click on Save.

## Appendix5

### 2D Image Analysis ( Measure )



1. Enclose interesting regions by ROI

Line on interesting positions by ROI

2. Click "measure" .

4. According to click "Measure All ROIs", then the information of all ROI is calculated on Region Measurement.

3. The information of ROI is calculated on Region Measurement.

5. The information of all ROIs

Measure	ROI No.	5	Statistics	CHS1	CHS2
CenterX	150.780	Integrate	121878548.000	54771708.000	
CenterY	79.732	Average	1244.509	559.277	
Area	6129.813	Max	4095.000	3227.000	
Perimeter	10.239	Min	1.000	1.000	
Range	3999.000	Range	3186.000		
StdDev	2175.309	StdDev	1518.002		

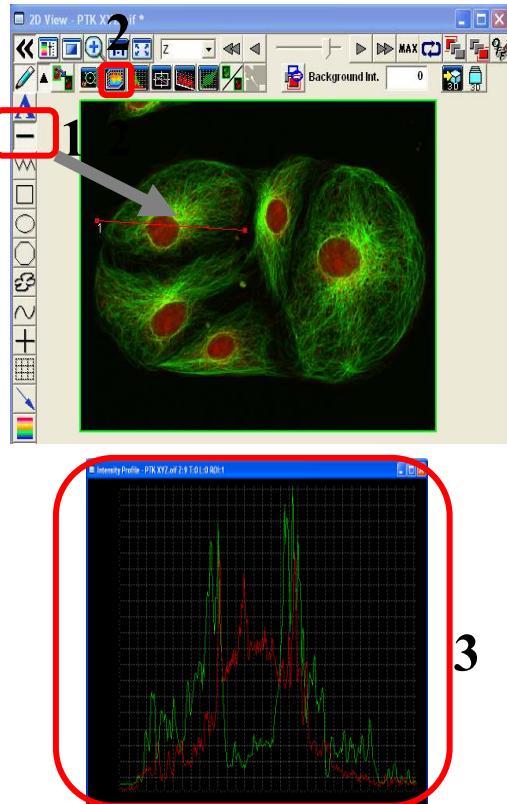
  

ROI	CenterX	CenterY	Area	Perimeter	Integration	Average	Max	Min	Range	StdDev	3StdDev	Integration	Average	Max	Min	Range	StdDev	3StdDev
1	57.171	49.438	3129.625	241.490	5478264.000	1107.926	4095.000	95.000	4000.000	710.261	2130.783	2952481.000	658.076	3590.000	28.000	3562.000	523.518	1567.554
2	112.522	53.402	1470.188	194.764	4620457.000	1301.724	4095.000	97.000	4001.000	883.602	2650.807	7837013.000	758.280	3590.000	28.000	3440.000	561.877	1685.630
3	51.900	87.103	3274.688	273.215	2573667.000	1003.410	4095.000	94.000	4001.000	700.397	2101.192	9839166.000	569.504	3415.000	53.000	3362.000	443.623	1330.869
4	80.180	111.524	1732.438	211.246	4386227.000	879.766	3836.000	83.000	3753.000	657.656	1972.967	7880740.000	645.072	3380.000	25.000	3355.000	523.061	1569.183
5	150.780	79.732	6120.813	313.258	1878548.000	1244.509	4095.000	96.000	3999.000	725.103	2175.309	4771708.000	559.277	3227.000	41.000	3186.000	439.334	1318.002

Count	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5
Average	90.511	76.240	3145.550	246.795	5987432.600	1107.467	4043.200	93.000	3950.200	735.404	2206.212	4656221.600	638.042	3416.000	35.000	3381.000	498.083	1494.2
Max	150.780	111.524	6120.813	313.258	1878548.000	1301.724	4095.000	97.000	4001.000	883.602	2650.807	4771708.000	758.280	3590.000	53.000	3562.000	561.877	1685.630
Min	51.900	49.438	1470.188	194.764	4386227.000	879.766	3836.000	83.000	3753.000	657.656	1972.967	7837013.000	559.277	3227.000	25.000	3186.000	439.334	1318.002
Range	98.879	62.087	4650.625	118.495	7492321.000	421.958	259.000	14.000	248.000	225.947	677.849	6934695.000	199.002	363.000	28.000	376.000	122.543	367.0
StdDev	41.309	25.569	1048.840	47.735	8699715.061	172.621	115.828	5.701	110.244	86.561	259.683	5124831.492	80.326	132.286	11.811	137.208	54.102	162.3
3StdDev	123.928	76.707	5546.521	143.205	6099145.184	517.864	347.485	17.103	330.731	259.683	779.050	5374494.476	240.979	396.857	35.433	411.624	162.305	486.5

## 2D Image Analysis ( Line Intensity Profile on the 2D image )

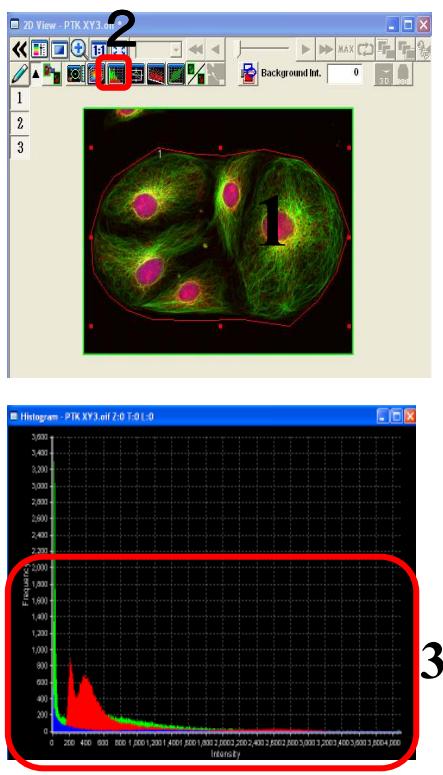


1. Line on the 2D image by ROI

2. Click "Intensity Profile,"

3. "Intensity Profile," on the line is shown as intensity graph .

## 2D Image Analysis ( Histogram )

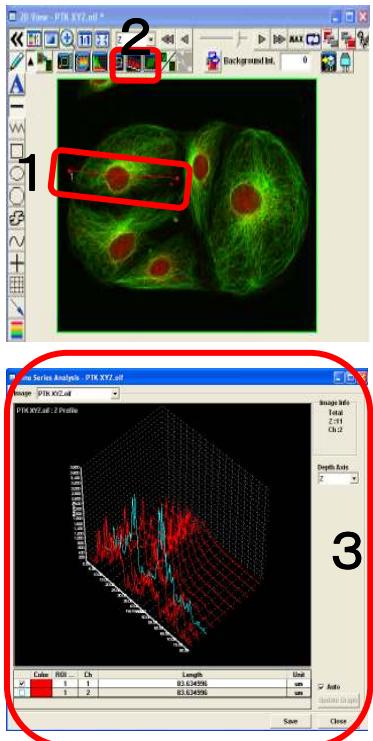


1. Enclose an interested area by ROI.

2. Click "Histogram,"

3. "Histogram," window is shown as a graph, frequency of intensity of each pixels is plotted on the area enclosed by ROI.

## 2D Image Analysis ( Line Series Analysis )

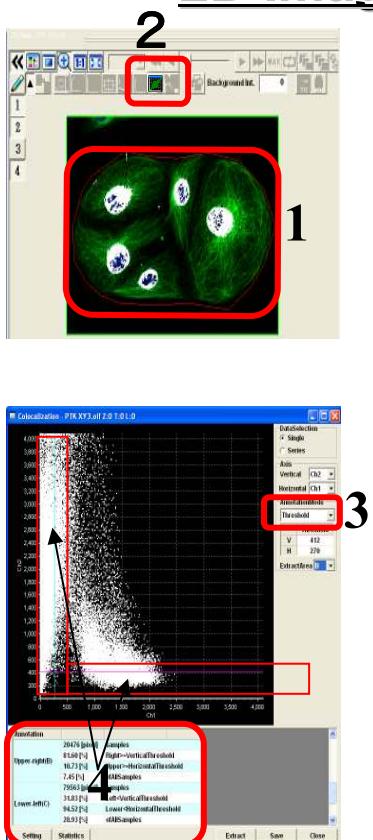


1. Line on the interesting region.

2. Click "Line Series Analysis,,

3. Intensity of Z position/ time on the line is shown as a graph .

## 2D Image Analysis ( Co-localization )



1. Enclose an interesting region by ROI.

2. Click

3. Select **Threshold** from Annotation Mode.

4. According to move Thresholds of X,Y axis to right and left ,ups and down **(Enclose red color X,Y axis)**, Co-localization result between 2ch is changed .

Information of Co-localization is listed under the scatter plot.



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